Variation in the human lipidome associated with coffee consumption as revealed by quantitative targeted metabolomics

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Abbreviations: KORA, Cooperative Health Research in the Region of Augsburg; MONICA, Monitoring of Trends and Determinants in Cardiovascular Disease; SM(OH), N-hydroxylacyloylsphingosyl-phosphocholine; SM(OH, COOH), N-hydroxyldicarboacyloylsphingosyl-phosphocholine

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ABSTRACT

The effect of coffee consumption on human health is still discussed controversially. Here we report results from a metabolomics study of coffee consumption, where we measured 363 metabolites in blood serum of 284 male participants of the KORA (Cooperative Health Research in the Region of Augsburg) study population, aged between 55 and 79 years. A statistical analysis of the association of metabolite concentrations and the number of cups of coffee consumed per day showed that coffee intake is positively associated with two classes of sphingomyelins, one containing a hydroxy-group (SM(OH)) and the other having an carboxy-group (SM(OH,COOH)). In contrast long- and medium-chain additional acylcarnitines were found to decrease with increasing coffee consumption. It is noteworthy that the concentration of total cholesterol also rises with an increased coffee intake in this study group. The here observed association between these hydroxylated and carboxylated sphingolipid species and coffee intake may be induced by changes in the cholesterol levels. Alternatively, these molecules may act as scavengers of oxidative species which decrease with higher coffee intake. In summary, we demonstrate strong positive associations between coffee consumption and two classes of sphingomyelins and a negative association between coffee consumption and long- and medium-chain acylcarnitines.

INTRODUCTION

Coffee is one of the most widely consumed beverages in the world [1]. Roasted, it consists of more than 800 components, including lipids, diterpenes such as cafestol and kahweol, carbohydrates, volatile and nonvolatile acids (e.g. acetic acid, chlorogenic acids), alkaloids such as caffeine, minerals and vitamins (e.g. niacin) [2]. The effects of coffee consumption on human health – beneficial as well as detrimental – are still discussed in the literature: Detrimental effects of coffee intake to health are thought to occur with respect to coronary heart disease: Most case-control studies suggest that the consumption of more than five cups of coffee per day leads to an increased risk for coronary heart disease or myocardial infarction [3-7]. However, the majority of the cohort studies could not confirm this observation [4, 8-12]. Also, the assumption that coffee induces hypertension is only supported by case-control studies [13-16], but not by cohort studies [8, 17-20]. The suspicion that coffee consumption may induce cancer could not be confirmed in several studies [21-23]. In contrast, a positive effect of coffee on cancer could be detected depending on the particular tissue: One metaanalysis and one review reporting the results of several case-control studies as well as of cohort studies show that, in the case of colorectal cancer, an inverse association between this cancer and coffee intake is observed and that, in contrast, cohort studies could not confirm such an association [24, 25]. However, for the hepatocellular carcinoma, the results of casecontrol as well as of cohort studies yielded a reduced risk of this cancer for heavy coffee drinkers [26-29].

Another potentially beneficial effect of coffee lies in the prevention of diabetes mellitus type 2. Higdon and Frei [8] reported that in six out of nine cohort studies a significant inverse association between coffee consumption and the risk for diabetes type 2 was found. For

example, a study with 1,000 Finnish twins found that the intake of more than seven cups of coffee per day reduces the risk of diabetes by 35% compared to a consumption of at most two cups per day [30]. An even larger effect, namely a decrease of 50%, was observed in a study with 17,000 Dutch individuals, again when more than seven cups of coffee per day were ingested [31]. Although an improvement of the glucose tolerance or of the insulin sensitivity by coffee consumption could not be determined in short-term trials, several cohort studies indicate that coffee consumption supports the maintenance of a normal glucose tolerance [32-35]. Coffee also seems to have a positive effect on the prevention of Parkinson's Disease [36-39]. Moreover, coffee consumption appears to lower the suicide risk [40-42] as well as the risk for dementia [43]. These studies demonstrate that coffee has many diverse effects on human health. However, it should be taken into account that some of these effects may also be explained by common, not yet identified covariates that may be traced back to general life style factors. Thus, more research in this field is needed.

In this study, we took a targeted quantitative metabolomics approach to detect coffee-induced changes on human metabolism. Targeted metabolomics aims at the quantitative measurement of ideally all key metabolites in a body fluid, thereby representing a snapshot of the full metabolic state of the organism. Changes in the metabolic profiles can be caused for example by specific food intake, disease or medication. Such changes can be detected in a very detailed manner using metabolomics techniques, so that the affected pathways may eventually be identified. This aim is now coming into reach with recent advances in the field of high resolution electrospray ionization tandem mass spectrometry. Here, we use this technology to determine the metabolic profiles of 284 participants from the KORA (Cooperative Health Research in the Region of Augsburg) study population [44]. Information on many different factors concerning life style (e.g. smoking and alcohol consumption), disease state or

medication use are available for each participant. These factors and their association with metabolic changes are presently analyzed in a number of separate studies which are based on this metabolomics dataset. Some of which have already been published for the association with nicotine consumption [45] and genetic variation [46]. The results from these studies also provide some kind of independent validation of this metabolomics dataset. In the present study we specifically examined the consequences of coffee consumption on the human metabolism.

MATERIAL & METHODS

Experimental Setup

KORA is a research platform for population-based surveys and subsequent follow-up studies in the fields of health economics, epidemiology, genetics and health care research [44]. It provides a multitude of different parameters, such as life style factors (nutrition, physical activity, smoking, alcohol consumption, etc.), sociodemographic variables, and medical history. The dataset presented here comes from the F3 study conducted in 2004-2005 as a follow-up of the third MONICA (Monitoring of Trends and Determinants in Cardiovascular Disease) survey (S3; 1994/1995). For this F3 study, 3006 of the 4856 participants of the S3 were reexamined for a second time, thus, 10 years apart from the baseline examination. From this group, 284 male individuals, aged between 55 and 79 years at the examination for F3, were chosen randomly for our analysis and were again recruited one to two years later for additional blood sampling. Out of this group, 239 of these individuals provided details concerning their coffee consumption (only participants younger than 75 were asked for this information). 82 participants (~34.3%) drank one to two cups of coffee per day, 100

individuals (~41.9%) had three to seven cups/day, twelve (~5.0%) regularly drank eight and more cups/day, and 45 (~18.8%) totally abstained from coffee consumption. As a control, information on the consumption of green tea and black tea was also analyzed. The study design was approved by an ethical committee and the informed consent of all participating subjects was obtained.

Blood samples

Blood samples for metabolic analysis were collected during 2006. To avoid variation due to circadian rhythm, blood was drawn in the morning between 8 and 10 am after a period of overnight fasting. Material was immediately horizontally shaken (10 min), followed by 40 min resting at 4°C to obtain complete coagulation. The material was then centrifuged (2000g; 4°C). Serum was aliquoted and stored for 2-4 hours at 4°C, after which it was deep frozen to -80°C until mass spectrometry analysis.

Metabolite profiling

Targeted metabolite profiling by electrospray ionization (ESI) tandem mass spectrometry (MS/MS) was performed at *Biocrates life sciences AG*, Austria. The technique is described in detail by patent US 2007/0004044 (accessible online at http://www.freepatentsonline.com/20070004044.html). A summary of the method can be found in ([47]; [48]). A comprehensive overview of the field and the related technologies is given in the review paper by Wenk [49]. Briefly, the assay preparation was done by an automated robotics system (Hamilton Robotics GmbH) on a special double-filter 96 well plate containing isotope labeled internal standards. Assays used 10µl plasma and include Phenylisothiocyanate (PITC)-derivatisation of amino acids, extraction with organic solvent and several pipetting, drying, and centrifugation steps. Flow injection analysis coupled with multiple reaction monitoring scans (FIA MS/MS) on a 4000 QTrap instrument (Applied Biosystems) was used for quantification of amino acids, acylcarnitines, sphingomyelins, phosphatidylcholines, and hexoses. LC-MS/MS methods using multiple reaction monitoring, neutral loss and precursor ion scans were applied for biogenic amines, eicosanoids and hydroxylated fatty acid derivatives as well as for intermediates of the energy metabolism. The quantification of the metabolites is achieved by reference to appropriate internal standards. The method is proven to be in conformance with 21CFR (Code of Federal Regulations) Part 11, which implies proof of reproducibility within a given error range. This measurement platform has been used in the past in different academic and industrial applications [45, 46, 50-52].

Metabolite spectrum

Concentrations of all analyzed metabolites are reported in µM. In total, 363 different metabolites were screened and detected in plasma: 18 amino acids, nine reducing mono-, diand oligosaccharides (abbreviated as Hn for n-hexose, dH for desoxyhexose, UA for uronic acid, HNAc for N-acetylglucosamine), 21 acylcarnitines (Cx:y, where x denotes the number of carbons in the side chain and y the number of double bonds), seven hydroxylacylcarnitines (C(OH)x:y) and dicarboxylacylcarnitines (Cx:y-DC), free carnitine (C0), seven biogenic amines, seven prostaglandins and 293 lipids. These lipids are subdivided into 14 different ceramides (Cer) and glucosylceramides (GlcCer), 71 different sphingomyelins (SMx:y) and sphingomyelin-derivatives, such as N-hydroxyldicarboacyloylsphingosyl-phosphocholine (SM(OH,COOH)x:y) and N-hydroxylacyloylsphingosyl-phosphocholine (SM (OH)x:y), five glycero-phosphatidic acids (PA), 124 glycero-phosphatidylcholines (PC), 42 glycerophosphatidylethanolamines (PE), four phosphatidylglycerols (PG), glycerophosphatidylinositols (PI) and glycero-phosphatidylinositol-bisphosphate (PIP2), and three glycero-phosphatidylserines (PS). Glycero-phospholipids are further differentiated with respect to the presence of ester (a) and ether (e) bonds in the glycerol moiety, where two letters (aa, ae, or ee) denote that the first as well as the second position of the glycerol unit are bound to a fatty acid residue, while a single letter (a or e) indicates a bond with only one fatty acid residue. E.g. PC ae 33:1 denotes a plasmalogen phosphatidylcholine with 33 carbons in the two fatty acid side chains and a single double bond in one of them. In some cases, the mapping of metabolite names to individual masses can be ambiguous. For example, stereochemical differences are not always discernable, neither are isobaric fragments.

Statistical analysis

The statistical analysis system R (http://www.r-project.org/) and SPSS for Windows (Version 16.0, Chicago: SPSS Inc.) were used for the statistical analysis. For the identification of metabolites influenced by coffee consumption, a Kendall's test was applied to each variable (metabolite concentration). Kendall's test is a non-parametric test that calculates the correlation of two variables based on the rank of their data. This test was chosen as it does neither require a linear correlation of the variables nor a normal distribution of the data. To control for the effect of testing multiple hypotheses, the positive false discovery rate (q-value) was computed, which is a measure for the fraction of false positives appearing even if the test itself was significant [53]. The rank correlation coefficient Kendall's tau was determined for every metabolite to indicate the direction of the correlation.

RESULTS

We analyzed the metabolic dataset to identify those metabolites that are most strongly associated with coffee consumption. The results of the Kendall tests are given in *Table 1*. Most remarkable is the association of two sphingomyelin classes (SM(OH,COOH)x:y and SM(OH)x:y) with coffee intake, with p-values ranging between 7.5x10⁻⁴ and 2.4x10⁻⁵. The Kendall q-value confirms a very low false positive rate (smaller than 2.5%). The positive values of the Kendall's tau for all sphingomyelins listed in Table 1 suggest that the sphingomyelins species SM(OH,COOH)x:y and SM(OH)x:y positively correlate with coffee intake. In order to further test this hypothesis, we calculated the sum of the concentrations of all SM(OH,COOH)x:y as well as the sum of the concentrations of all SM(OH)x:y and applied the Kendall's test to each. The sums of these sphingomyelin classes also correlate positively with coffee consumption, as shown in Figures 1 and 2.

Besides the sphingomyelins, also long- and medium-chain acylcarnitines showed low p-values concerning the association to the coffee intake (*Tab.1*; *Supplemental Tab.1*). However, this association concerns mainly the two groups of coffee drinkers consuming 1-2 cups/day and 3-7 cups/day respectively. For the individual long- and medium-chain acylcarnitines as well as for the sum of all acylcarnitines with a chain length of at least 10, the Kendall's tau yielded negative values. Thus, the coffee consumption and the plasma concentration of long- and medium-chain acylcarnitines are inversely associated.

From the remaining metabolites none was associated with coffee consumption with a positive False Discovery Rate (pFDR; q-value) lower than 5%. As a positive control, showing that null findings within this rest of the metabolites cannot be attributed to a potentially poor data quality, we note the identification of a number of very significant associations in other studies concerning different metabolites from this same metabolomics dataset, such as different

phosphatidyl-choline, -ethanolamine and -inositol species, as well as amino acids and short and medium chain length acyl carnitines, with p-values down to 10^{-21} as documented by [46] and [45]. As a negative control, the effect of green tea as well as that of black tea on the presented set of metabolites was analyzed. For neither kind of tea a significant correlation with any metabolite could be found, especially all q-values were larger than 0.1. This fact emphasizes the significance of the observed associations between the coffee consumption and the two reported sphingomyelin species.

DISCUSSION

In this paper, we analyzed the influence of coffee consumption on the metabolism of 239 male participants of the KORA project. The measurement of 363 metabolites by tandem mass spectrometry provides a deeper insight into changes in the metabolic state caused by coffee intake. Humans are exposed to many different environmental influences, such as various nutrition, physical activity or medication. Therefore, it is remarkable that three groups of metabolites clearly associated with coffee consumption could be identified in this study. Among these metabolites are the long- and medium-chain acylcarnitines, whose concentrations decrease with rising coffee consumption. One explanation might be the effect of niacin, which is a component of coffee [54]. Niacin lowers the concentration of triglycerides and free fatty acids in the plasma [55-59]. A reduced concentration of these triglycerides and free fatty acids causes a decrease of the concentration of acylcarnitines, which are synthesized during the transport of fatty acids into the mitochondrium for β-oxidation. Thus, an increase of fat storage could be a consequence.

The sphingomyelin classes SM(OH,COOH)x:y and SM(OH)x:y showed a positive association to coffee intake. The synthesis of sphingomyelin is located in the golgi apparatus and is based on the transfer of the phosphorylcholine from a phosphatidylcholine to a ceramide [60, 61]. A ceramide consists of a sphingosine and a fatty acid that is added to the C2 of the sphingosine by N-acetylation [62]. SM(OH)x:y is formed by adding an αhydroxylated fatty acid [63]. For SM(OH,COOH)x:y this fatty acid additionally carries a carboxyl group, presumably at the ω-position (Figure 3). In contrast to SM(OH)x:y, the sphingomyelin class SM(OH,COOH)x:y is not described in much detail in the literature. In principle, there are two mechanistic explanations for the biogenic formation of this molecule: Either an unsaturated fatty acid chain in a sphingomyelin is oxidized, or an existing αhydroxy-1, ω -dicarboxyl fatty acid is added to the C2 position of the sphingosine. α -hydroxy-1,ω-dicarboxyl fatty acids have been found in the cell wall of Legionella in a study by Sonesson et al. [64]. An alternative explanation for the formation of SM(OH,COOH)x:y might be an oxidative attack on a double bond of the fatty acid moiety. During this process the carboxyl group might be built at the ω-position. Hence, SM(OH,COOH)x:y could function as a scavenger and might therefore reduce the risk of oxidative damage leading for instance to cancer. Sargis and Subbaiah [65] reported that sphingomyelins are able to protect cholesterol from oxidation, but the exact mechanism is still unknown and requires further investigation.

Sphingomyelins are mainly located in biological membranes, including the myelin sheath of nerve cell axons. These membranes are mainly composed of different lipids, proteins and further components such as cholesterol, which interacts with phospholipids to stabilize them and thus supports their ability to build membranes. Among the phospholipids sphingomyelins show the strongest interaction with cholesterol [66, 67]. Moreover the analysis of Gronberg *et*

al. [67] showed that cholesterol increases the packing of SM(OH)x:y in the membrane. Both, SM(OH,COOH)x:y and SM(OH)x:y, were positively associated with coffee consumption in our study. This could be linked to an increased cholesterol level in plasma. It is known, that cholesterol stabilizes and increases the packing of sphingomyelins and, hence, supports the formation of membranes. In course of the KORA project, blood parameters, including cholesterol, have been determined for each participant. Our analysis showed that total cholesterol increases with coffee consumption in this study group (Kendall p-value: 1.1·10⁻⁴; Kendall's tau: 0.165, Figure 4). Using data from the entire F3-population indicates that this positive association is still significant after adjustment for age and sex (data not shown). Based on this observation, and the described function of cholesterol, one may, thus, speculate that the increased cholesterol levels and the increased SM(OH,COOH)x:y and SM(OH)x:y are functionally related.

Whether the intake of coffee really increases the concentration of cholesterol is still discussed in literature. In the 1980s, evidence was provided that coffee consumption causes a higher concentration of cholesterol in blood [68]. But the subsequent studies found that this effect is based mainly on the diterpenes cafestol and kahweol, which are only present in unfiltered coffee [69-71] Thus, this would indicate that there is no association between coffee consumption and cholesterol. This aspect is important, since the people in southern Germany (location of the KORA project) mainly drink filtered coffee. Exactly the opposite result, a positive association of filtered coffee intake with the concentration of cholesterol in blood, was reported by the studies of [72], [73], [74] and [75]. So we can conclude that there are indicators for this positive correlation between filtered coffee and cholesterol, but still more evidence is needed.

An additional aspect worth considering is the association of sphingomyelin with atherosclerosis. Since decades it is known, that sphingomyelin accumulates in atheromas [76] Recent studies have confirmed the association of atherosclerosis with an increased sphingomyelin level in animals as well as in humans [77-80]. In this context one might speculate that coffee consumption heightens the risk of atherosclerosis by increasing the levels of the two resulting classes of sphingomyelins.

Concerning these sphingomyelins, an interesting association was identified in a genome-wide association study. In this study, the above discussed (coffee-associated) sphingomyelin species are found to be strongly associated with a genetic variant (SNP) in the PLEK gene. The PLEK gene codes for pleckstrin, a protein that has been proposed to facilitate protein/lipid interactions and to affect membrane structure. It is supposed to enable the localization to the cell membrane by acting as a signal adapter molecule. Ma *et al.* [81] report, that pleckstrin associates with human platelet membranes and supports the formation of membrane projections from transfected (Cos-1) cells. Pleckstrin is recruited into natural killer cell membranes, when these cells are activated [82], and it associates with the membranes of stimulated macrophages [83]. This observation raises the question, whether coffee consumption and variation in the PLEK gene may have an interacting effect on the level of the described sphingomyelins, and possibly related medical outcomes.

Our results demonstrate that the quantitative measurement of more than 360 metabolites by tandem mass spectrometry may detect unexpected correlations in human metabolism. Especially the influence of coffee on the two classes of sphingomyelins is a remarkable result of our analysis. This could be a starting point for further research on the function of these sphingomyelins.

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AUTHOR DISCLOSURES

K.M.W. is an employee of Biocrates Life Sciences AG. This private company offers products and services in the field of targeted quantitative metabolomics research. The other authors have nothing to declare.

REFERENCES

- [1] International Coffee Organization, Annual review. 2006/2007.
- [2] Spiller, M. A., in: Spiller, G. A. (Ed.), Caffeine, CRC Press, Boca Raton 1998, pp. 97-161.
- [3] Greenland, S., A meta-analysis of coffee, myocardial infarction, and coronary death. *Epidemiology* 1993, 4, 366-374.
- [4] Kawachi, I., Colditz, G. A., Stone, C. B., Does coffee drinking increase the risk of coronary heart disease? Results from a meta-analysis. *Br Heart J* 1994, 72, 269-275.
- [5] Hammar, N., Andersson, T., Alfredsson, L., Reuterwall, C., *et al.*, Association of boiled and filtered coffee with incidence of first nonfatal myocardial infarction: the SHEEP and the VHEEP study. *J Intern Med* 2003, *253*, 653-659.
- [6] Palmer, J. R., Rosenberg, L., Rao, R. S., Shapiro, S., Coffee consumption and myocardial infarction in women. *Am J Epidemiol* 1995, *141*, 724-731.
- [7] Tavani, A., Bertuzzi, M., Negri, E., Sorbara, L., La Vecchia, C., Alcohol, smoking, coffee and risk of non-fatal acute myocardial infarction in Italy. *Eur J Epidemiol* 2001, *17*, 1131-1137.
- [8] Higdon, J. V., Frei, B., Coffee and health: a review of recent human research. *Crit Rev Food Sci Nutr* 2006, *46*, 101-123.
- [9] Myers, M. G., Basinski, A., Coffee and coronary heart disease. *Arch Intern Med* 1992, 152, 1767-1772.
- [10] Willett, W. C., Stampfer, M. J., Manson, J. E., Colditz, G. A., *et al.*, Coffee consumption and coronary heart disease in women. A ten-year follow-up. *JAMA* 1996, *275*, 458-462.
- [11] Woodward, M., Tunstall-Pedoe, H., Coffee and tea consumption in the Scottish Heart Health Study follow up: conflicting relations with coronary risk factors, coronary disease, and all cause mortality. *J Epidemiol Community Health* 1999, *53*, 481-487.

- [12] Kleemola, P., Jousilahti, P., Pietinen, P., Vartiainen, E., Tuomilehto, J., Coffee consumption and the risk of coronary heart disease and death. *Arch Intern Med* 2000, *160*, 3393-3400.
- [13] Jee, S. H., He, J., Whelton, P. K., Suh, I., Klag, M. J., The effect of chronic coffee drinking on blood pressure: a meta-analysis of controlled clinical trials. *Hypertension* 1999, 33, 647-652.
- [14] Nurminen, M. L., Niittynen, L., Korpela, R., Vapaatalo, H., Coffee, caffeine and blood pressure: a critical review. *Eur J Clin Nutr* 1999, *53*, 831-839.
- [15] James, J. E., Critical review of dietary caffeine and blood pressure: a relationship that should be taken more seriously. *Psychosom Med* 2004, *66*, 63-71.
- [16] Noordzij, M., Uiterwaal, C. S., Arends, L. R., Kok, F. J., *et al.*, Blood pressure response to chronic intake of coffee and caffeine: a meta-analysis of randomized controlled trials. *J Hypertens* 2005, *23*, 921-928.
- [17] Dawber, T. R., Kannel, W. B., Gordon, T., Coffee and cardiovascular disease. Observations from the framingham study. *N Engl J Med* 1974, *291*, 871-874.
- [18] Bertrand, C. A., Pomper, I., Hillman, G., Duffy, J. C., Micheli, I., No relation between coffee and blood pressure. *N Engl J Med* 1978, *299*, 315-316.
- [19] Lancaster, T., Muir, J., Silagy, C., The effects of coffee on serum lipids and blood pressure in a UK population. *J R Soc Med* 1994, *87*, 506-507.
- [20] Salvaggio, A., Periti, M., Miano, L., Zambelli, C., Association between habitual coffee consumption and blood pressure levels. *J Hypertens* 1990, *8*, 585-590.
- [21] Leviton, A., Methylxanthine consumption and the risk of ovarian malignancy. *Cancer Lett* 1990, *51*, 91-101.
- [22] Tavani, A., La Vecchia, C., Coffee and cancer: a review of epidemiological studies, 1990-1999. *Eur J Cancer Prev* 2000, *9*, 241-256.

- [23] Zeegers, M. P., Kellen, E., Buntinx, F., van den Brandt, P. A., The association between smoking, beverage consumption, diet and bladder cancer: a systematic literature review. *World J Urol* 2004, *21*, 392-401.
- [24] Giovannucci, E., Meta-analysis of coffee consumption and risk of colorectal cancer. *Am J Epidemiol* 1998, *147*, 1043-1052.
- [25] Tavani, A., La Vecchia, C., Coffee, decaffeinated coffee, tea and cancer of the colon and rectum: a review of epidemiological studies, 1990-2003. *Cancer Causes Control* 2004, *15*, 743-757.
- [26] Gelatti, U., Covolo, L., Franceschini, M., Pirali, F., *et al.*, Coffee consumption reduces the risk of hepatocellular carcinoma independently of its aetiology: a case-control study. *J Hepatol* 2005, *42*, 528-534.
- [27] Gallus, S., Bertuzzi, M., Tavani, A., Bosetti, C., et al., Does coffee protect against hepatocellular carcinoma? *Br J Cancer* 2002, 87, 956-959.
- [28] Inoue, M., Yoshimi, I., Sobue, T., Tsugane, S., Influence of coffee drinking on subsequent risk of hepatocellular carcinoma: a prospective study in Japan. *J Natl Cancer Inst* 2005, *97*, 293-300.
- [29] Shimazu, T., Tsubono, Y., Kuriyama, S., Ohmori, K., *et al.*, Coffee consumption and the risk of primary liver cancer: pooled analysis of two prospective studies in Japan. *Int J Cancer* 2005, *116*, 150-154.
- [30] Carlsson, S., Hammar, N., Grill, V., Kaprio, J., Coffee consumption and risk of type 2 diabetes in Finnish twins. *Int J Epidemiol* 2004, *33*, 616-617.
- [31] van Dam, R. M., Feskens, E. J., Coffee consumption and risk of type 2 diabetes mellitus. *Lancet* 2002, *360*, 1477-1478.
- [32] Yamaji, T., Mizoue, T., Tabata, S., Ogawa, S., *et al.*, Coffee consumption and glucose tolerance status in middle-aged Japanese men. *Diabetologia* 2004, *47*, 2145-2151.

- [33] Soriguer, F., Rojo-Martinez, G., de Antonio, I. E., Coffee consumption and type 2 diabetes mellitus. *Ann Intern Med* 2004, *141*, 321-323; author reply 323-324.
- [34] Agardh, E. E., Carlsson, S., Ahlbom, A., Efendic, S., *et al.*, Coffee consumption, type 2 diabetes and impaired glucose tolerance in Swedish men and women. *J Intern Med* 2004, *255*, 645-652.
- [35] van Dam, R. M., Dekker, J. M., Nijpels, G., Stehouwer, C. D., *et al.*, Coffee consumption and incidence of impaired fasting glucose, impaired glucose tolerance, and type 2 diabetes: the Hoorn Study. *Diabetologia* 2004, *47*, 2152-2159.
- [36] Ascherio, A., Zhang, S. M., Hernan, M. A., Kawachi, I., *et al.*, Prospective study of caffeine consumption and risk of Parkinson's disease in men and women. *Ann Neurol* 2001, 50, 56-63.
- [37] Ascherio, A., Chen, H., Schwarzschild, M. A., Zhang, S. M., *et al.*, Caffeine, postmenopausal estrogen, and risk of Parkinson's disease. *Neurology* 2003, *60*, 790-795.
- [38] Ascherio, A., Weisskopf, M. G., O'Reilly, E. J., McCullough, M. L., *et al.*, Coffee consumption, gender, and Parkinson's disease mortality in the cancer prevention study II cohort: the modifying effects of estrogen. *Am J Epidemiol* 2004, *160*, 977-984.
- [39] Ross, G. W., Abbott, R. D., Petrovitch, H., Morens, D. M., *et al.*, Association of coffee and caffeine intake with the risk of Parkinson disease. *JAMA* 2000, *283*, 2674-2679.
- [40] Klatsky, A. L., Armstrong, M. A., Friedman, G. D., Coffee, tea, and mortality. *Ann Epidemiol* 1993, *3*, 375-381.
- [41] Kawachi, I., Willett, W. C., Colditz, G. A., Stampfer, M. J., Speizer, F. E., A prospective study of coffee drinking and suicide in women. *Arch Intern Med* 1996, *156*, 521-525.
- [42] Tanskanen, A., Tuomilehto, J., Viinamaki, H., Vartiainen, E., et al., Heavy coffee drinking and the risk of suicide. Eur J Epidemiol 2000, 16, 789-791.

- [43] Eskelinen, M. H., Ngandu, T., Tuomilehto, J., Soininen, H., Kivipelto, M., Midlife coffee and tea drinking and the risk of late-life dementia: a population-based CAIDE study. *J Alzheimers Dis* 2009, *16*, 85-91.
- [44] Holle, R., Happich, M., Löwel, H., Wichmann, H. E., KORA A Research Platform for Population Based Health Research *Das Gesundheitswesen* 2005, *S* 01, 19-25.
- [45] Wang-Sattler, R., Yu, Y., Mittelstrass, K., Lattka, E., *et al.*, Metabolic profiling reveals distinct variations linked to nicotine consumption in humans--first results from the KORA study. *PLoS ONE* 2008, *3*, e3863.
- [46] Gieger, C., Geistlinger, L., Altmaier, E., Hrabe de Angelis, M., *et al.*, Genetics meets metabolomics: a genome-wide association study of metabolite profiles in human serum. *PLoS Genet* 2008, *4*, e1000282.
- [47] Weinberger, K. M., Graber, A., Using Comprehensive Metabolomics to Identify Novel Biomarkers. *Screening Trends in Drug Discovery* 2005, *6*, 42–45.
- [48] Weinberger, K. M., [Metabolomics in diagnosing metabolic diseases]. *Ther Umsch* 2008, 65, 487-491.
- [49] Wenk, M. R., The emerging field of lipidomics. Nat Rev Drug Discov 2005, 4, 594-610.
- [50] Butler, A., Schneider, R., Floyd, E., Roffi, C., et al., Metabonomic profiling to identify biomarkers of vascular injury in canines and humans. *Proc Cambridge Healthtech Institute's* 7th Annual Meeting, Metabolic Markers for Drug Development and Clinical Studies, Orlando, FL, December 4-5 2006.
- [51] Bernardo, K., Stoeggl, W., Marini, G., Zaccaria, P., et al., A fully automated sample preparation and electrospray ionisation tandem mass spectrometry based approach to identify and quantify lipids an application to monitor lipid changes in cardiovascular disease patients. Proc 17th International Mass Spectrometry Conference, Prague, Czech Republic, August 25 September 1 2006.

- [52] Altmaier, E., Ramsay, S. L., Graber, A., Mewes, H. W., *et al.*, Bioinformatics analysis of targeted metabolomics--uncovering old and new tales of diabetic mice under medication. *Endocrinology* 2008, *149*, 3478-3489.
- [53] Storey, J. D., The positive false discovery rate: A Bayesian interpretation and the q-value. *The Annals of Statistics* 2003, *31*, 2013–2035.
- [54] Casal, S., Oliveira, M. B., Alves, M. R., Ferreira, M. A., Discriminate analysis of roasted coffee varieties for trigonelline, nicotinic acid, and caffeine content. *J Agric Food Chem* 2000, 48, 3420-3424.
- [55] Carlson, L. A., Oro, L., The effect of nicotinic acid on the plasma free fatty acid; demonstration of a metabolic type of sympathicolysis. *Acta Med Scand* 1962, *172*, 641-645.
- [56] Grundy, S. M., Mok, H. Y., Zech, L., Berman, M., Influence of nicotinic acid on metabolism of cholesterol and triglycerides in man. *J Lipid Res* 1981, *22*, 24-36.
- [57] DiPalma, J. R., Thayer, W. S., Use of niacin as a drug. *Annu Rev Nutr* 1991, *11*, 169-187.
 [58] Wahlberg, G., Walldius, G., Effects of nicotinic acid treatment on fatty acid composition of plasma lipids and adipose tissue in hyperlipidaemia. *Scand J Clin Lab Invest* 1992, *52*, 547-553.
- [59] Hwang, Y. C., Bakr, S., Ramasamy, R., Bergmann, S. R., Relative importance of enhanced glucose uptake versus attenuation of long-chain acyl carnitines in protecting ischemic myocardium. *Coron Artery Dis* 2002, *13*, 313-318.
- [60] Futerman, A. H., Stieger, B., Hubbard, A. L., Pagano, R. E., Sphingomyelin synthesis in rat liver occurs predominantly at the cis and medial cisternae of the Golgi apparatus. *J Biol Chem* 1990, 265, 8650-8657.
- [61] Pagano, R. E., What is the fate of diacylglycerol produced at the Golgi apparatus? *Trends Biochem Sci* 1988, *13*, 202-205.

- [62] Goni, F. M., Contreras, F. X., Montes, L. R., Sot, J., Alonso, A., Biophysics (and sociology) of ceramides. *Biochem Soc Symp* 2005, 177-188.
- [63] Uchida, Y., Hara, M., Nishio, H., Sidransky, E., *et al.*, Epidermal sphingomyelins are precursors for selected stratum corneum ceramides. *J Lipid Res* 2000, *41*, 2071-2082.
- [64] Sonesson, A., Moll, H., Jantzen, E., Zahringer, U., Long-chain alpha-hydroxy-(omega-1)-oxo fatty acids and alpha-hydroxy-1,omega-dioic fatty acids are cell wall constituents of Legionella (L. jordanis, L. maceachernii and L. micdadei). *FEMS Microbiol Lett* 1993, *106*, 315-320.
- [65] Sargis, R. M., Subbaiah, P. V., Protection of membrane cholesterol by sphingomyelin against free radical-mediated oxidation. *Free Radic Biol Med* 2006, *40*, 2092-2102.
- [66] Sankaram, M. B., Thompson, T. E., Interaction of cholesterol with various glycerophospholipids and sphingomyelin. *Biochemistry* 1990, *29*, 10670-10675.
- [67] Gronberg, L., Ruan, Z. S., Bittman, R., Slotte, J. P., Interaction of cholesterol with synthetic sphingomyelin derivatives in mixed monolayers. *Biochemistry* 1991, *30*, 10746-10754.
- [68] Thelle, D. S., Arnesen, E., Forde, O. H., The Tromso heart study. Does coffee raise serum cholesterol? *N Engl J Med* 1983, *308*, 1454-1457.
- [69] Zock, P. L., Katan, M. B., Merkus, M. P., van Dusseldorp, M., Harryvan, J. L., Effect of a lipid-rich fraction from boiled coffee on serum cholesterol. *Lancet* 1990, *335*, 1235-1237.
- [70] Ahola, I., Jauhiainen, M., Aro, A., The hypercholesterolaemic factor in boiled coffee is retained by a paper filter. *J Intern Med* 1991, *230*, 293-297.
- [71] Urgert, R., Katan, M. B., The cholesterol-raising factor from coffee beans. *Annu Rev Nutr* 1997, 17, 305-324.

- [72] Superko, H. R., Bortz, W., Jr., Williams, P. T., Albers, J. J., Wood, P. D., Caffeinated and decaffeinated coffee effects on plasma lipoprotein cholesterol, apolipoproteins, and lipase activity: a controlled, randomized trial. *Am J Clin Nutr* 1991, *54*, 599-605.
- [73] Fried, R. E., Levine, D. M., Kwiterovich, P. O., Diamond, E. L., *et al.*, The effect of filtered-coffee consumption on plasma lipid levels. Results of a randomized clinical trial. *JAMA* 1992, *267*, 811-815.
- [74] Christensen, B., Mosdol, A., Retterstol, L., Landaas, S., Thelle, D. S., Abstention from filtered coffee reduces the concentrations of plasma homocysteine and serum cholesterol--a randomized controlled trial. *Am J Clin Nutr* 2001, 74, 302-307.
- [75] Strandhagen, E., Thelle, D. S., Filtered coffee raises serum cholesterol: results from a controlled study. *Eur J Clin Nutr* 2003, *57*, 1164-1168.
- [76] Smith, E. B., Intimal and medial lipids in human aortas. *Lancet* 1960, 1, 799-803.
- [77] Park, T. S., Panek, R. L., Rekhter, M. D., Mueller, S. B., *et al.*, Modulation of lipoprotein metabolism by inhibition of sphingomyelin synthesis in ApoE knockout mice. *Atherosclerosis* 2006, *189*, 264-272.
- [78] Jiang, X. C., Paultre, F., Pearson, T. A., Reed, R. G., et al., Plasma sphingomyelin level as a risk factor for coronary artery disease. *Arterioscler Thromb Vasc Biol* 2000, 20, 2614-2618.
- [79] Nelson, J. C., Jiang, X. C., Tabas, I., Tall, A., Shea, S., Plasma sphingomyelin and subclinical atherosclerosis: findings from the multi-ethnic study of atherosclerosis. *Am J Epidemiol* 2006, *163*, 903-912.
- [80] Liu, J., Zhang, H., Li, Z., Hailemariam, T. K., et al., Sphingomyelin Synthase 2 Is One of the Determinants for Plasma and Liver Sphingomyelin Levels in Mice. *Arterioscler Thromb Vasc Biol* 2009.

- [81] Ma, A. D., Brass, L. F., Abrams, C. S., Pleckstrin associates with plasma membranes and induces the formation of membrane projections: requirements for phosphorylation and the NH2-terminal PH domain. *J Cell Biol* 1997, *136*, 1071-1079.
- [82] al-Aoukaty, A., Rolstad, B., Maghazachi, A. A., Recruitment of pleckstrin and phosphoinositide 3-kinase gamma into the cell membranes, and their association with G beta gamma after activation of NK cells with chemokines. *J Immunol* 1999, *162*, 3249-3255.
- [83] Brumell, J. H., Howard, J. C., Craig, K., Grinstein, S., *et al.*, Expression of the protein kinase C substrate pleckstrin in macrophages: association with phagosomal membranes. *J Immunol* 1999, *163*, 3388-3395.

FIGURES AND TABLES

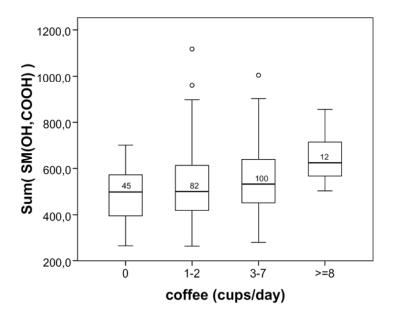


FIGURE 1: Boxplots of the plasma concentrations [μ M] of the sum of all sphingomyelin species SM(OH,COOH)x:y as a function of coffee consumption. The Kendall p-value for the association is 4.9×10^{-5} . Boxes extend from 1^{st} quartile (Q_1) to 3^{rd} quartile (Q_3); median is indicated as a horizontal line; whiskers are drawn to the observation that is closest to, but not more than a distance of $1.5(Q_3-Q_1)$ from the end of the box. Observations that are more distant than this are shown individually on the plot. The number of individuals in each group is given in the boxes.

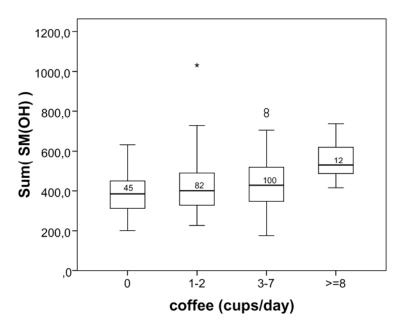


FIGURE 2: Boxplots for the sphingomyelin species SM(OH)x:y (legend see Fig. 1). The Kendall p-value for the association is $1.0x10^{-4}$.

FIGURE 3: Assumed structure of the sphingomyelin species SM(OH)x:y (A) and SM(OH,COOH)x:y (B).

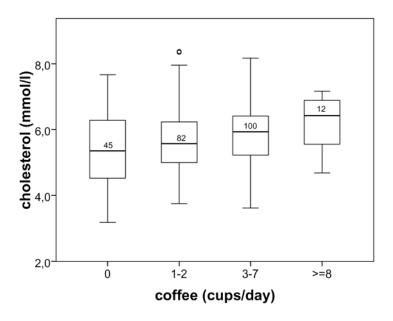


FIGURE 4: Boxplots of the plasma concentrations of total cholesterol levels (legend see Fig. 1). The Kendall p-value for the association is 1.1×10^{-4} .

^{*} can also be unsaturated

Table 1: Results of the Kendall rank test, with p- and q-value as well as the Kendall's tau; all tested metabolites yielding a Kendall p-value smaller than 10^{-3} and a q-value smaller than $3x10^{-2}$ are listed. The sums of SM(OH,COOH)x:y, of SM(OH)x:y and of all acylcarnitines Cx:y with chain length greater or equal to ten are indicated in italics. In cases where alternative assignments of the metabolites are possible, these are indicated by a '*'.

metabolites	mean	standard	Kendall	Kendall	Kendall's
		deviation	p-value	q-value	tau
	μΜ	μΜ			
SM (OH,COOH) 20:2	64.741	22.394	2.4E-05	3.1E-03	0.179
SM (OH,COOH) 16:2 *	84.938	26.854	2.6E-05	3.2E-03	0.179
SM (OH) 20:3	172.874	53.820	4.2E-05	4.2E-03	0.174
C16:1	0.036	0.014	4.3E-05	4.2E-03	-0.174
Sum(SM(OH,COOH)x:y)	528.757	149.989	4.9E-05	2.2E-03	0.172
C10:1	0.166	0.075	9.8E-05	6.8E-03	-0.165
Sum(SM(OH)x:y)	423.809	126.783	1.0E-04	3.0E-03	0.165
SM (OH,COOH) 18:2	92.451	30.334	1.1E-04	7.3E-03	0.164
C12:1	0.122	0.054	1.3E-04	7.9E-03	-0.163
C14:1	0.126	0.064	1.4E-04	8.3E-03	-0.162
SM (OH) 22:1	44.567	17.038	1.4E-04	8.5E-03	0.162
C14:2	0.052	0.024	1.7E-04	9.9E-03	-0.159
Sum(Cx10:y)	1.463	0.510	1.9E-04	3.3E-03	-0.159
SM (OH,COOH) 24:0	20.928	8.206	2.2E-04	1.2E-02	0.157
SM (OH,COOH) 18:1	19.272	6.467	4.0E-04	1.7E-02	0.150
SM (OH) 28:0	59.325	22.334	5.0E-04	1.9E-02	0.148
C6	0.089	0.036	7.5E-04	2.5E-02	-0.143